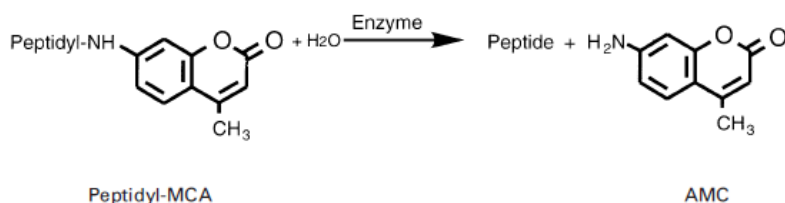


# Assay Methods Using Peptidyl-MCA Substrates (1)

## Principle



The initial rate of increase in the AMC concentration can be monitored 1) fluorometrically at  $\lambda_{ex} = 380 \text{ nm}$  and  $\lambda_{em} = 460 \text{ nm}$  (Fig. 1a) or 2) photometrically at 370 nm (Fig. 1b).

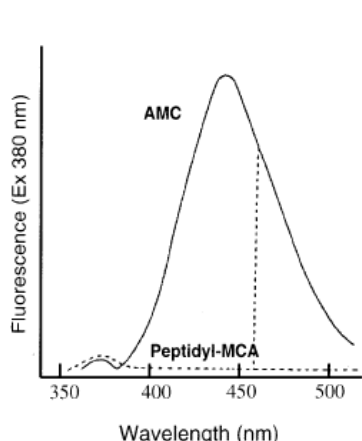


Fig. 1a Fluorescence Spectra

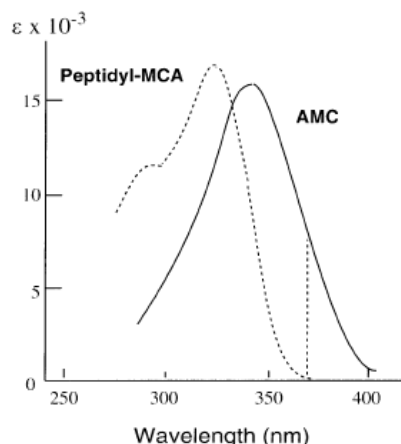


Fig. 1b UV-Absorption Spectra

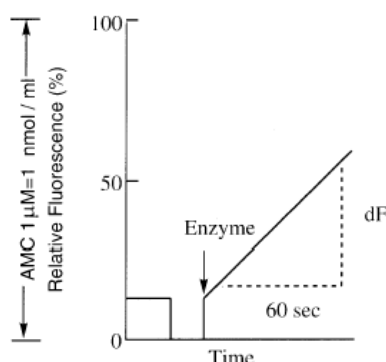
## Reagents

- 1) Substrate stock solution: Vial, in DMSO at 10 mM
- 2) AMC stock solution: Content of vial (Code 3099-v AMC), in DMSO at 1 mM
- 3) Buffer
- 4) Enzyme solution

## Procedure

Choose the proper conditions for the measurement, such as substrate concentration and sensitivity setting, depending on the purpose of the experiment and the instrument available. Described here is one of the recommended procedures for the fluorometric method (initial-rate method).

- 1) Set a fluorescence spectrophotometer at  $\lambda_{ex} = 380 \text{ nm}$  and  $\lambda_{em} = 460 \text{ nm}$  at 25 °C (1.0 Relative fluorescence unit at  $10^{-6} \text{ M}$  of AMC)
- 2) Pipette 2940  $\mu\text{l}$  of buffer and 30  $\mu\text{l}$  of substrate stock solution into the cuvette
- 3) Incubate in the fluorescence spectrophotometer for 3-4 min (for temperature equilibration)
- 4) Add 30  $\mu\text{l}$  of enzyme solution
- 5) Record the increase of the fluorescence intensity for 3-4 min
- 6) Calculate the amount of AMC released using the following equation



Amount (nmol) of AMC released/min

$$\begin{aligned}
 &= \frac{1 \text{ nmol} \times dF\% \times 3 \text{ ml}}{1 \text{ ml} \times 100\% \times 1 \text{ min}} \\
 &= 0.03 \times dF \text{ nmol/min}
 \end{aligned}$$

\* Photometric measurement can be carried out by the same procedure as that of the fluorometric method using a UV spectrophotometer. Set the wavelength at 370 nm ( $\epsilon$  7700).

## Assay Methods Using Peptidyl-MCA Substrates (2)

### (Measurement on an Auto-Fluorescence Spectrophotometer for Multiplate)

#### Reagents

- 1) Substrate stock solution: Content of vial, in DMSO at 10 mM
- 2) AMC standard solution: Content of vial (Code 3099-v AMC), in DMSO at 1 mM
- 3) Buffer
- 4) Enzyme solution

#### Instrument: Auto-fluorescence spectrophotometer

Selection of filter:

380 nm, 390 nm or 355 nm filter will be recommended for measurement.

In using 355 nm filter, observe caution for overlapping of fluorescence of the substrate itself.

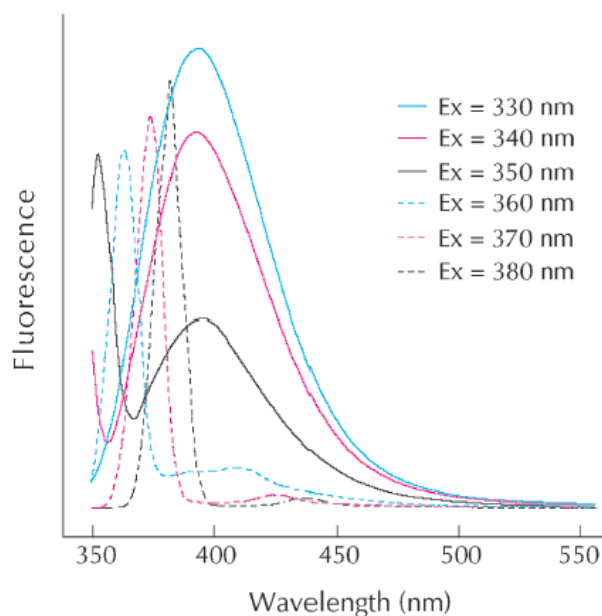


Fig. 2a Fluorescence Spectra of Peptidyl-MCA (Substrate)

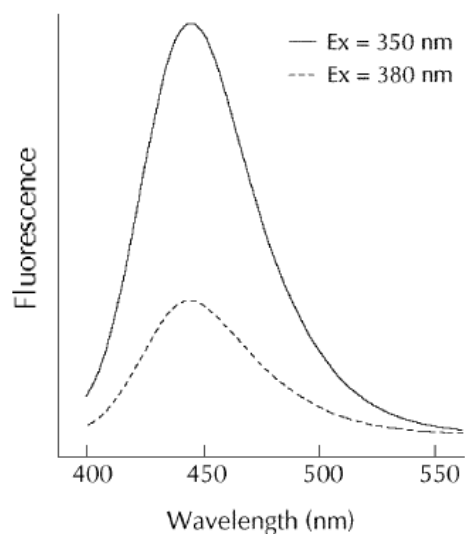


Fig. 2b Fluorescence Spectra of AMC (Product)

#### Procedure

Choose the proper conditions for the measurement, such as substrate, enzyme concentration and other reaction conditions, depending on the purpose of the experiment.

- 1) Set the auto-fluorescence spectrophotometer at  $\lambda_{ex} = 380 \text{ nm}$ ,  $\lambda_{em} = 460 \text{ nm}$  at  $25^\circ\text{C}$   
(1.0 Relative fluorescence unit at  $10^{-6} \text{ M}$  of AMC)
  - 2) Pipette  $160 \mu\text{l}$  of buffer and  $20 \mu\text{l}$  of substrate solution in well for final concentration of  $100 \mu\text{M}$
  - 3) Incubate the plate in the fluorescence spectrophotometer for 3-4 min (for temperature equilibration)
  - 4) Take the multiplate out and add  $20 \mu\text{l}$  of enzyme solution in each well
  - 5) Mount the plate in the fluorescence spectrophotometer
  - 6) Record the increase in fluorescence intensity for 30 min with a premixing time of 3 sec
  - 7) Calculate the amount of released AMC
-

## Assay Methods Using Peptidyl-MCA Substrates (3)

(Example for Caspase-7 using an Auto-fluorescence Spectrophotometer for Multiplate)

### Reagents

- 1) Substrate stock solution: Content of vial (Code 3171-v Ac-DEVD-MCA), in DMSO at 10 mM
- 2) AMC standard solution: Content of vial (Code 3099-v AMC), in DMSO at 1 mM
- 3) Buffer: ICE standard buffer (100 mM HEPES-KOH, pH 7.5, 10% sucrose (w/v), 0.1% CHAPS (w/v) , 10 mM DTT, 0.1 mg/ml ovalbumin)
- 4) Enzyme solution: rec Caspase-7 in buffer

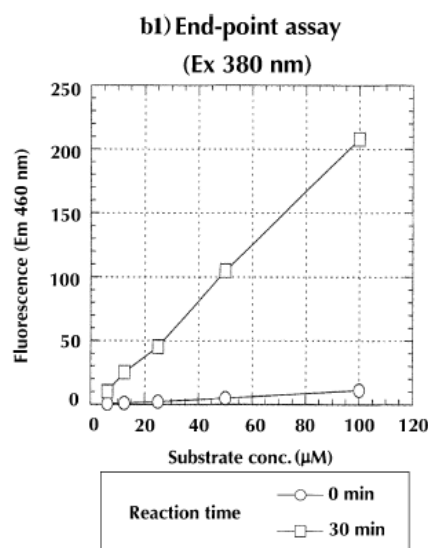
### Insurument: Fluoroskan Ascent (Labsystems)

This instrument can be used for both initial rate assay and end point assay

### Procedure

- a) Initial rate assay for 30 min at  $\lambda_{ex} = 390 \text{ nm}$ ,  $\lambda_{em} = 460 \text{ nm}$
- b) End-point (30 min) assay at 5 different substrate concentrations at  $\lambda_{ex} = 355 \text{ nm}$  or  $380 \text{ nm}$ ,  $\lambda_{em} = 460 \text{ nm}$

- 1) Set a fluorescence spectrophotometer at  $\lambda_{ex} = 390 \text{ nm}$  ( or  $380 \text{ nm}$ ,  $355 \text{ nm}$ ),  $\lambda_{em} = 460 \text{ nm}$ .  
Relative fluorescence is determined with  $10^{-6} \text{ M}$  of AMC at each condition
- 2) Substrate: Dilute the stock solution with the buffer ( $\times 10$ ,  $\times 20$ ,  $\times 40$ ,  $\times 80$ ,  $\times 160$ )
- 3) Caspase-7: Dissolve in buffer
- 4) Pipette  $160 \mu\text{l}$  of buffer and  $20 \mu\text{l}$  of substrate solutions (1, 1/2, 1/4, 1/8, 1/16 mM) in each well
- 5) Incubate in the fluorescence spectrophotometer for 3-4 min for temperature equilibration
- 6) Take the multiplate out and add  $20 \mu\text{l}$  of enzyme solution to each well
- 7) Mount the plate in the fluorescence spectrophotometer
- 8) Calculate the amount of released AMC



### Results

